

# Hybridization, Introgression, and the Nature of Species Boundaries

RICHARD G. HARRISON AND ERICA L. LARSON

From the Department of Ecology and Evolutionary Biology, Cornell University, Corson Hall, Ithaca, NY 14853 (Harrison); and the Division of Biological Sciences, University of Montana, Missoula, MT 59812 (Larson).

Address correspondence to Richard G. Harrison at the address above, or e-mail: [rgh4@cornell.edu](mailto:rgh4@cornell.edu).

---

## Abstract

Species can be defined as populations that are diagnosably distinct, reproductively isolated, cohesive, or exclusive groups of organisms. Boundaries between species in sympatry are maintained by intrinsic barriers to gene exchange; these boundaries may not be uniform in space, in time, or across the genome. Here, we explore the nature of the species boundary, defined as the phenotypes/genes/genome regions that remain differentiated in the face of potential hybridization and introgression. We emphasize that species boundaries are semipermeable, with permeability (gene exchange) being a function of genome region. The early evidence for semipermeable species boundaries came from data on differential introgression in hybrid zones. This “genic view” of species was common in the hybrid zone literature even when few molecular markers were available to characterize genome-wide patterns of variation. Now, molecular tools allow detailed characterization of differentiation between diverging lineages and patterns of variation across natural hybrid zones, but the questions being asked by evolutionary biologists have remained much the same. Recent data (from DNA sequences and genotypes) reinforce earlier conclusions about the semipermeable nature of most species boundaries. However, debate persists over the nature and extent of genome divergence that accompanies speciation.

**Subject areas:** *Population structure and phylogeography*

**Key words:** *genomic divergence, hybrid zones, reproductive isolation, speciation*

---

Evolutionary and systematic biologists have regularly engaged in prolonged and sometimes acrimonious debates about species concepts and definitions; species have been variously defined as entities that are diagnosably distinct, reproductively isolated, cohesive, or exclusive (monophyletic) groups of organisms. Species concepts that focus on the importance of reproductive isolation (e.g., the Biological Species Concept [BSC] of Mayr and Dobzhansky) have often occupied center stage (Harrison 1998). The BSC has certainly provided the framework for the many empirical studies of speciation that have involved identifying the phenotypes and genotypes responsible for intrinsic barriers to gene exchange (e.g., Coyne and Orr 2004).

But the focus on reproductive isolation has frequently been questioned, and critics have particularly taken aim at the writings of Ernst Mayr. For example, Mallet (2008a) argued that Mayr, by emphasizing discontinuity and complete reproductive isolation, rejected Charles Darwin’s (correct) vision of continuity between varieties and species (Darwin 1859) and failed to acknowledge that divergence between lineages can be maintained in the face of gene flow. Similarly, Wu (2001), championing what he termed a “genic view” of species, suggested that Mayr’s BSC necessarily implies that reproductive

isolation is a “whole-genome concept” and “lose[s] its logical robustness” if we acknowledge that the extent of isolation varies across the genome. According to Wu (2001), the BSC must be a whole-genome concept because Mayr (1963) argued that the genotype is coadapted, or as Mayr (1942) put it, “a ‘physiological team.’”

Critics of the “isolation view” of species often seem to imply that most students of speciation in the late 20th century were devout disciples of Mayr and that those who defended the BSC also must have been believers in coadapted gene complexes and the sanctity of allopatric speciation. In fact, many evolutionary biologists in that era recognized continuity in degree of divergence, viewed reproductive isolation and barriers to gene exchange as potentially incomplete, and argued that species boundaries can be semipermeable. A “genic view” had already been proposed long before Wu’s (2001) article (more on this below). Even Mayr, condemned by Mallet for leading evolutionary biologists astray, apparently softened his views in later years. In *The Growth of Biological Thought* (1982), he discussed hybridization and wrote: “...for it seems as if some part of the genotype of the 2 species is not affected by the hybridization. The 2 species, in such a case, seem to remain “reproductively isolated,” in the sense

that they do not fuse into a single population, in spite of the leakage of certain of their genes.” (p. 285).

Here, we explore the nature of species boundaries and the importance of hybridization and introgression in defining such boundaries. We examine the notion that species boundaries are semipermeable, with permeability (gene exchange) being a function of genome region. This idea is not new and was widely discussed in the hybrid zone literature long before evolutionary biologists had access to the array of molecular markers that now allow characterization of genome-wide patterns of divergence. Beginning with the application of allozyme and mtDNA data to the study of hybrid zones (see [Harrison 1990](#) for a review), documenting patterns of differential introgression provided strong evidence for the semipermeable nature of species boundaries. The recent introduction of high-throughput sequencing allows characterization of patterns of variation and divergence for multiple markers across the genome; far more detailed views of the species boundary are now becoming available. It is, therefore, timely to examine the history of ideas and data that are relevant to the concept of species boundaries.

## Hybridization and Introgression

Natural hybridization can be defined as the interbreeding of individuals from 2 distinct populations or groups of populations. Individuals in those populations must be distinguishable on the basis of one or more heritable characters ([Harrison 1990, 1993](#)). Natural hybridization is most easily recognized when previously allopatric populations come together in secondary contact. Renewed sympatry often results in a hybrid zone, with parental types,  $F_1$  hybrids, and multiple generation hybrids and backcrosses present in varying proportions. The presence of diverse genotypes, the product of many generations of recombination, potentially allows fine-scale mapping of genes that contribute to reproductive isolation and estimates of selection on individual alleles ([Barton and Hewitt 1985; Barton and Gale 1993](#)). Thus, natural hybrid zones provide data not easily obtained from laboratory crosses because such crosses usually involve relatively few generations of recombination.

Introgression (or “introgressive hybridization”) describes the incorporation (usually via hybridization and backcrossing) of alleles from one entity (species) into the gene pool of a second, divergent entity (species) ([Anderson and Hubricht 1938; Anderson 1949](#)). Introgression is a relative term; alleles at one locus introgress with respect to alleles at other loci. That is, for the above definition to be applicable, some portion of the gene pool of each of the hybridizing taxa must remain constant and uncontaminated such that we can actually recognize that 2 distinct gene pools exist. As we will discuss below, the genes that define the 2 gene pools and make them distinct are those that comprise the species boundary.

Differential introgression, a phenomenon documented in many hybrid zones, refers to the observation that alleles at some loci introgress more than others. In theory, globally advantageous alleles will tend to introgress easily (“adaptive

introgression”; e.g., see [Whitney et al. 2006; Pardo-Diaz et al. 2012; Hedrick 2013](#)); neutral alleles will introgress to varying extents, but linkage to genes that contribute to local adaptation or reproductive isolation will inhibit their movement ([Barton 1979](#)). Alleles will introgress little or not at all when they represent variants at loci subject to divergent directional selection and/or loci that determine speciation phenotypes (phenotypes that are responsible for reproductive isolation; see [Shaw and Mullen 2011](#)). Thus, patterns of differential introgression across hybrid zones potentially allow identification of genes or genome regions that are important for local adaptation and speciation ([Payseur 2010; Nachman and Payseur 2012](#)).

The geographic pattern and spatial scale of introgression will depend on many factors, including the environmental context in which hybridization occurs, how far individuals disperse, and the nature of natural selection (e.g., contrast clinal “tension zones” with mosaic hybrid zones embedded in a patchy environment). Some authors (e.g., [Heiser 1973](#)) have differentiated between localized and dispersed introgression, distinguished by whether introgressed alleles are found only where the 2 parental types occur together (and hybridize) or whether alleles of one species flow into otherwise pure populations of the “other” species that may be geographically far from a hybrid zone.

## Species Boundaries

The term “species boundary” has been used frequently in the evolutionary biology literature. Introgression is often described as occurring “across species boundaries” (e.g., the introduction of techniques for mtDNA restriction fragment length polymorphism (RFLP) analysis led to a spate of papers that discussed mtDNA gene flow across the species boundary; [Ferris et al. 1983; Powell 1983; Harrison et al. 1987](#)). Recent articles refer to resolving, delimiting, or mapping species boundaries ([Bouck et al. 2005; Lemmon et al. 2007; Roe and Sperling 2007; Wagner et al. 2013](#)). However, exactly what the species boundary represents is not always made clear. It is certainly the case that the boundary in some way reflects the fact that gene flow between species is limited or prevented in nature by a set of intrinsic barriers. These barriers reflect phenotypic differences between species that impact whether individuals mate assortatively, whether after mating (or spawning or pollen release) gametes get together to form zygotes, or whether the zygotes thus formed give rise to viable and fertile adults.

The term species boundary can be used to refer to the geographic boundary between parapatric taxa. Although of obvious importance for understanding the ecology and recent history of the taxa, spatial boundaries are not what students of speciation mean when they use the term “species boundaries.” In many cases, delimiting or resolving species boundaries refers to “boundaries” that might be visualized in tree space. For species with relatively old divergence times, a phylogenetic approach can give straightforward results, where species boundaries are defined by the presence of exclusive or reciprocally monophyletic groups (although allopatric monophyletic groupings may not be recognized as species).

However, for pairs or groups of species that are products of recent divergence, or that continue to exchange genes, species boundaries can be difficult to define not only because there is little differentiation but also because there may be discordance among character sets or among different gene trees. Discordance can reflect differential introgression (see below) but is also expected because of ancestral polymorphism, random lineage sorting, and the long time required for many or most loci to achieve reciprocal monophyly (e.g., see Hudson and Coyne 2002). Discordance among individual gene trees has been documented in many different pairs or groups of species (Beltran et al. 2002; Machado and Hey 2003; Dopman et al. 2005; Putnam et al. 2007; Andrés et al. 2008; Nachman and Payseur 2012). In many cases, a provisional set of boundaries can be defined on the basis of phenotype (morphological, behavioral, or ecological traits). If a significant fraction of gene trees are concordant with the provisional tree, then those markers are often assumed to mark the species boundary, with discordant trees explained by shared ancestral polymorphism or ongoing gene exchange. In the most problematic cases, increasing the amount of molecular data can lead to resolution. Thus, Wagner et al. (2013) demonstrate that high-throughput DNA sequence data from restriction site-associated DNA (RAD) markers provide “unprecedented resolution of species boundaries” in Lake Victoria cichlid fish, a group for which previous phylogenetic analyses had consistently revealed extensive allele sharing between putative morphs or species.

Phylogenetic approaches assume that species should be exclusive or monophyletic groups, at least for some part of the genome. These approaches work equally well for taxa that are allopatric and those that are sympatric or parapatric. However, patterns of exclusivity for allopatric taxa provide no guarantee that these taxa would remain distinct in sympatry; exclusivity can arise simply as a product of geographic isolation over time (e.g., due to genetic drift), without necessarily impacting the potential for gene exchange when taxa become sympatric. In contrast, exclusive phylogenetic relationships for taxa that occur together (either broadly sympatric or in narrow hybrid zones) suggest that the taxa are indeed distinct species, in the sense that gene flow between them does not lead to fusion or homogenization. In these situations, the focus is on what maintains species boundaries. Mayr (1963) clearly recognized the importance of geographic context in defining species. He emphasized what he called a “nondimensional” species concept, “characterized by the non-interbreeding of 2 coexisting demes, uncomplicated by the dimensions of space and time.” (Mayr 1963, p. 669).

Thus, we might consider the species boundary to be defined by the phenotypes/genes/genome regions (or some subset thereof) that remain differentiated in the face of potential hybridization and introgression (i.e., when the entities in question are locally sympatric). This definition acknowledges that species boundaries do not necessarily extend across the entire genome, that alleles at some (perhaps many) loci can be exchanged between species, that species boundaries are semipermeable or porous, and that species boundaries can vary geographically. The words “or some subset thereof” are

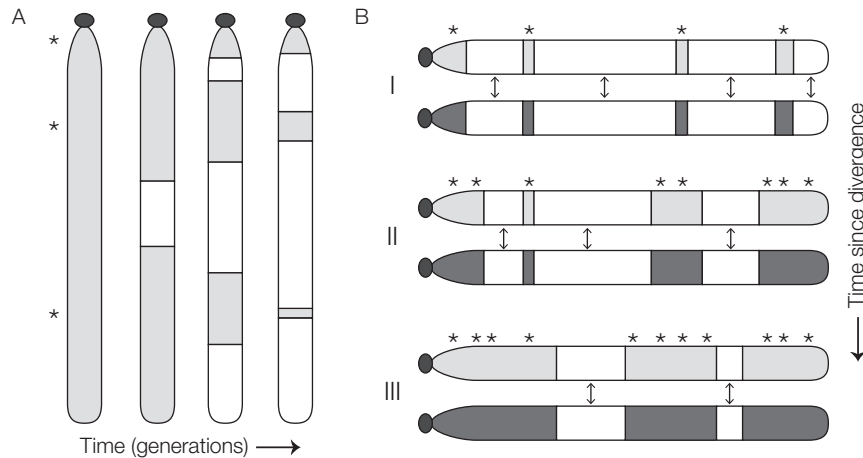
included because not all genome regions that remain distinct when taxa are sympatric necessarily contribute to reproductive isolation. In an extreme case, allelic differences at a single locus could result in perfect positive assortative mating or hybrid lethality and would prevent gene exchange across the entire genome; yet, the “species boundary” might be thought of as defined by a single locus.

## Semipermeable Species Boundaries

The “genic view” of species advocated by Wu (2001) was not a new idea, but his review article brought the idea to the attention of a larger community. The notion that gene flow and reproductive isolation are characteristics of genome regions, not entire genomes, was already well established in the hybrid zone literature in the 1980s. Key (1968, p. 19), in discussing hybridization in Morabine grasshoppers, wrote: “Thus the tension zones act like semipermeable membranes, holding back some genes and chromosomal rearrangements to varying degrees, but permitting others rather free passage.” And Bazykin (1969), commenting on models of sympatric speciation, introduced the concept of “isolation for part of the gene pool.” In an early review of hybrid zones, Barton and Hewitt (1981, p. 119), citing Bazykin 1969, wrote that “Strict application of the biological species concept might lead to different results for different loci; perhaps one can only define ‘groups of actually or potentially interbreeding natural populations’ ... at the gene level.” A figure in that review shows an example of how sequences on a chromosome become homogenized over time by gene flow, except in small regions surrounding genes subject to divergent directional selection (Figure 1). In a subsequent review of hybrid zones, Harrison (1990, pp. 98–99) wrote that “Boundaries [between species] are, therefore, semi-permeable, the permeability depending on the genetic marker .... Genetic isolation must be considered as a property of individual genes (or chromosome segments), not as a characteristic of entire genome.”

Some authors prefer to characterize the species boundary as “porous” rather than semipermeable. Porous is a synonym of permeable and means “easily crossed or penetrated” ([www.thefreedictionary.com/](http://www.thefreedictionary.com/)) or “easy to pass or get through” ([www.merriam-webster.com/dictionary/](http://www.merriam-webster.com/dictionary/)). In cell biology, semipermeable is a term used to describe membranes that are selective in allowing only certain molecules or ions to pass through. A semipermeable boundary between species implies that differential introgression is the result of a selective process, with alleles at some loci able to cross the boundary, whereas alleles at other loci cannot. The term porous does not imply selectivity.

The early evidence for semipermeable species boundaries came primarily from data on patterns of differential introgression across hybrid zones. In the 1980s, available molecular markers were few (allozyme markers were first used in evolutionary biology in 1966, mtDNA data [RFLPs] first appeared in 1979). Nonetheless, comparisons using those markers, together with observations of morphological or



**Figure 1.** Classic illustrations of the semipermeable nature of species boundaries. **(A)** Gene flow following secondary contact as depicted by [Barton and Hewitt \(1981\)](#). The vertical bars represent a chromosome with 3 loci contributing to reproductive barriers (indicated by \*). Immediately after contact, linkage disequilibrium along the chromosome will be high, but over time recombination breaks down associations among loci. Barrier genes or genes under divergent selection will remain differentiated (light gray regions), whereas alleles at loci that are neutral (white regions) will be exchanged between species. Many generations of recombination in hybrid zones allow fine-scale mapping of genes contributing to reproductive isolation and estimates of the strength of selection on individual alleles. **(B)** The idea that the genomes of diverging lineages become less permeable over time, as shown by [Wu \(2001\)](#). Each pair of horizontal bars represents chromosomes of 2 diverging lineages. Very recently diverged species (pair I) may have few genes contributing to reproductive isolation (indicated by \*). These regions will remain differentiated (represented by light and dark gray regions), whereas gene exchange can occur in other parts of the genome (white regions). With increasing genetic divergence (chromosome pairs II and III), an increasing number of loci contribute to reproductive barriers, thus restricting gene flow for a greater proportion of the genome.

behavioral traits, supported the view that some markers (or sorts of markers) introgress further/faster than others (Table 1 in [Harrison 1990](#)). The observation that the extent of introgression of mtDNA markers was often greater than that of nuclear encoded markers was explained by the fact that mtDNA sequences are unlinked to the nuclear genome and, therefore, unlinked to genes that contribute to reproductive isolation ([Barton and Jones 1983](#); [Harrison 1989](#)).

By combining many variable molecular markers (random amplified polymorphic DNAs) with a linkage map of those markers, [Rieseberg et al. \(1999\)](#) were able to identify chromosomal segments with reduced introgression across 3 replicate hybrid zones between 2 sunflower species (genus *Helianthus*). The consistent patterns for the 3 presumably independent hybrid zones strongly suggested that reduced introgression was the product of deterministic forces (i.e., selection), and indeed, many of the chromosomal blocks with reduced introgression were shown to be associated with hybrid pollen sterility (an important barrier to gene exchange in *Helianthus*). This article provided the first detailed analysis of differential introgression in the context of a genetic map and remains a classic in the hybrid zone literature.

Given the rapid advances in DNA sequencing and genotyping technology, patterns of differentiation and introgression for multiple markers can now be assayed relatively easily, even for organisms that lack substantial genomic resources. Methods for estimating the extent of introgression and for interpreting observed patterns have similarly made important

advances ([Gompert and Buerkle 2009, 2011, 2012](#); [Payseur 2010](#); [Fitzpatrick 2013](#)). These methods can be divided into 2 categories: 1) those that analyze geographic clines (how allele and genotype frequencies change over space) and 2) those that employ genomic clines, in which changes in genotype frequencies for individual loci are examined “along a genomic admixture gradient” ([Gompert and Buerkle 2009](#), p. 1207). Both approaches can define patterns of differential introgression, but a genomic cline approach is particularly useful in mosaic hybrid zones, where it may not be possible to define a simple geographic transect, except at very fine spatial scales.

## The Genetic and Genomic Architecture of Species Boundaries

The “genetic architecture of species boundaries” refers to the number, effect size, and chromosomal distribution of the genes that encode phenotypes that result in barriers to gene exchange (speciation phenotypes). Discussions of genetic and genomic architectures have proliferated in recent years, as new and more efficient DNA sequencing and genotyping technologies have emerged. Comparisons between individuals from sister species or from races/strains/subspecies that are in the early stages of divergence can now be made at the level of whole-genome sequences, sequences from targeted regions of reduced complexity (e.g., transcriptome sequences

or RAD sequences), or for hundreds to hundreds of thousands of single-nucleotide polymorphisms (SNPs).

One of the early articles to make such a comparison, between 2 forms (now named species) of mosquitoes in the genus *Anopheles*, found that divergence appeared to be restricted to 3 regions of the genome and labeled these regions “genomic islands of speciation” (Turner et al. 2005). The term “genomic islands” stuck, and the subsequent literature has elaborated on the geographic/topographic imagery, although in some cases “islands of differentiation” has replaced “islands of speciation” (e.g., Nosil et al. 2009). Invoking similar imagery, some have suggested that a more common pattern may be “archipelagoes” or “continents” of speciation (Michel et al. 2010). It is not clear that the imagery of oceans, sea level, and terrestrial topography provides a useful context for discussing genetic architecture (Harrison 2012). Indeed, simply identifying regions that are significantly elevated in divergence remains a challenge and depends on (often unstated) assumptions about historical demography.

We are interested in how and where genes that determine speciation phenotypes are arrayed on chromosomes. We are also interested in how selection influences allele frequencies at these loci and in the impact of that selection on surrounding chromosome regions. The expected size of genome regions that remain differentiated in the face of some gene flow will depend on selection and recombination. It will also depend on the frequency of individuals heterozygous for population-specific markers (positive assortative mating will reduce this frequency) and on the reproductive success of those individuals. It is only in such individuals that recombination between population-specific alleles can occur. Via and West (2008) coined the term “divergence hitchhiking” to describe the fact that when there is the potential for hybridization between diverging populations, divergent selection and nonrandom mating reduce effective recombination rates (from those expected based simply on map distance). Whether “divergence hitchhiking” should result in larger islands of differentiation, as claimed by Via and West (2008), remains controversial (Nosil et al. 2009; Via 2009, 2012; Feder and Nosil 2010; Feder et al. 2012; Flaxman et al. 2012).

Many recent studies have characterized genome-wide patterns of divergence between closely related species. Table 1 summarizes data collected from a diversity of taxa. These studies represent a range of approaches for surveying patterns of variation across the genome and for identifying regions that exhibit excess divergence. However, the majority of studies have relied on estimates of  $F_{ST}$  (e.g., an “ $F_{ST}$  outlier” approach; Beaumont and Balding 2004) or other relative measures of divergence. Because relative measures of divergence (including  $F_{ST}$ ) depend both on divergence between and variation within populations, elevated  $F_{ST}$  can be due to reduced nucleotide diversity within populations (Charlesworth 1998; Nachman and Payseur 2012; Cruickshank and Hahn 2014). As a consequence, high  $F_{ST}$  values could reflect loss of variation within populations (e.g., as a result of a selective sweep), rather than excess divergence. It is, therefore, worth revisiting case histories for which  $F_{ST}$  outliers have been identified to determine the cause of

elevated  $F_{ST}$  (see Cruickshank and Hahn 2014). Future studies need to include comparisons based on absolute measures of sequence divergence.

Genome-wide comparisons between recently diverged forms or species suggest that divergence is not restricted to a few discrete regions. Several studies that compare ecologically distinct but morphologically indistinguishable forms claim that there is widespread, but heterogeneous, divergence across the genome. These comparisons include the M and S forms of *Anopheles gambiae* (Lawniczak et al. 2010) and host races of *Rhagoletis pomonella* (Michel et al. 2010). Similarly, divergences between hybridizing flycatchers (*Ficedula*) and between oceanic and freshwater threespine sticklebacks are also highly heterogeneous across the genome, with many “divergence islands” (Hohenlohe et al. 2010; Ellegren et al. 2012). However, exactly what is meant by “widespread” or “heterogeneous” divergence remains unclear. Indeed, few generalizations are yet possible; this is not surprising, given the very recent development of efficient sequencing/genotyping methods, the importance of having a reference genome or a dense linkage map, and the additional difficulty of making comparisons when very different sets of markers and analyses have been used for different pairs of taxa (Table 1). Among the clear patterns that have emerged are the observations that differentiation is greater in regions of low recombination (Nachman and Payseur 2012) and very often on sex chromosomes (Carneiro et al. 2010; Lawniczak et al. 2010; Ellegren et al. 2013). In fact, a number of models predict the accumulation of barrier genes in regions of restricted recombination (e.g., within inversions, adjacent to centromeres) (Noor et al. 2001; Rieseberg 2001; Navarro and Barton 2003).

Genome-wide comparisons (ranging from modest numbers of microsatellite loci to full genome sequences) also can provide important insights into the evolutionary history of recent speciation/diversification events. An increasing number of studies have revealed evidence of hybridization among diverging lineages (Patterson et al. 2006; Putnam et al. 2007; Grant and Grant 2010; Garrigan et al. 2012; Cui et al. 2013; Keller et al. 2013; Nadeau et al. 2013; The Heliconius Genome Consortium 2012; Prüfer et al. 2014). These observations are consistent with a variety of scenarios for diversification in the face of at least episodic gene flow and lend support to the notion that hybridization allows introgression of adaptive traits (see below) and can, in some cases, lead to the origin of novel traits.

Although comparisons between recently diverged allopatric lineages can document the genomic landscape of genetic differentiation between species, observed differences between allopatric populations may or may not persist if populations come into contact. In contrast, hybrid zones allow us to examine directly the maintenance of genetic differentiation between sympatric or parapatric taxa. Often the product of secondary contact between forms that have been allopatric for at least some of their recent history (Barton and Hewitt 1985; Harrison 1990), natural hybrid zones provide direct information on patterns of differential introgression. In most study systems, alleles at some loci introgress

**Table 1** Studies of genome-wide patterns of differentiation between recently diverged pairs of taxa that are isolated by reproductive barriers

Taxa compared	Organism	Time	Barriers	Populations (sample size)	Markers (library type)	$F_{ST}$	Outliers	References
<i>Acyrtosiphon pisum</i> “alfalfa,” “red clover,” and “pea”	Pea aphid	0.008–0.016 Ma	Habitat isolation	3 host plants from 3 regions (N = 180)	390 microsatellites (whole genome)	0.069–0.17 <sup>a</sup>	2.8%	Jaquéry et al. (2012), also Via and Hawthorne (2001), Via and West (2008), Smadja et al. (2012), and Via et al. (2012)
<i>Anopheles gambiae</i> “M,” “S,” and “Bamako”	Mosquito	—	Habitat isolation, assortative mating	1 sympatric (N = 60)	400 000 SNPs (whole genome)	—	—	Lawniczak et al. (2010), Neafsey et al. (2010) also Turner et al. (2005), Turner and Hahn (2007), White et al. (2010), and Weetman et al. (2011)
<i>Coregonus clupeaformis</i> “normal” and “dwarf”	Lake whitefish	0.06 Ma	Habitat isolation	1 sympatric (N = 24)	2203 SNPs (exon capture)	0.046	12%	Hebert et al. (2013), also Campbell and Bernatchez (2004), Renaud et al. (2010, 2011), and Gagnaire et al. (2013)
<i>Ficedula albicollis</i> and <i>Ficedula hypoleuca</i>	Flycatcher	>2 Ma	Assortative mating, hybrid sterility	2 allopatric (N = 20)	3.81M SNPs (whole genome)	0.357	2.7%	Ellegren et al. (2013)
<i>Gasterosteus aculeatus</i> “freshwater,” “oceanic,” “benthic,” and “limnetic”	Stickleback	0.012 Ma	Habitat isolation, assortative mating	34 allopatric (N = 196)	1159 SNPs (EST)	0.193 (0.031–0.383)	4.0% <sup>b</sup>	Jones, Grubherr, et al. (2012), also Hohenlohe et al. (2010, 2012) and Jones, Chan, et al. (2012)
<i>Gasterosteus aculeatus</i> “lake” and “stream”	Stickleback	0.012 Ma	Habitat isolation, assortative mating	8 allopatric (N = 216)	4127–8417 SNPs (RADseq)	0–0.149 <sup>a</sup>	—	Roesli et al. (2012), also Deagle et al. (2012)
<i>Gryllus firmus</i> and <i>Gryllus pennsylvanicus</i>	Field cricket	0.2 Ma	Habitat isolation, assortative mating, postmating prezygotic	2 allopatric (N = 30)	9731 SNPs (RNAseq)	—	—	Andrés et al. (2013)
<i>Helianthus petiolaris</i> “dune” and “non-dune”	Sunflower	0.01 Ma	Habitat isolation	20 allopatric (N = 100)	19 539 SNPs (RADseq)	0.121	1.7%	Andrew and Rieseberg (2013)
<i>Helianthus annuus</i> and <i>H. petiolaris</i>	Sunflower	1.8 Ma	Habitat isolation, hybrid sterility	10 allopatric (N = 20)	27 994 SNPs (RADseq)	0.316	—	Andrew and Rieseberg (2013), also Yatabe et al. (2007), Strausberg et al. (2009), Gompert and Buerkle (2009), and Kane et al. (2009)
<i>Helianthus annuus</i> ssp. <i>annuus</i> , <i>H. annuus</i> ssp. <i>texasanus</i> and <i>Helianthus debilis</i>	Sunflower	—	Habitat isolation, hybrid sterility	13 allopatric (N = 378)	88 microsatellites	—	3.4%	Scascitelli et al. (2010)
<i>Heliconius melpomene</i> ssp., <i>Heliconius timareta</i> ssp., <i>Heliconius heurippa</i> , <i>Heliconius cydno</i> ssp., and <i>Heliconius becale</i>	Butterfly	—	Assortative mating, hybrid inviability	5 sympatric, 2 parapatric (N = 60)	4078 SNPs (RADseq)	—	7.0%	Nadeau et al. (2013) also Nadeau et al. (2012) and The Heliconius Genome Consortium (2013)

**Table 1** Continued

Taxa compared	Organism	Time	Barriers	Populations (sample size)	Markers (library type)	$F_{ST}$	Outliers	References
<i>Honea behmoreana</i> and <i>Honea forsteriana</i>	Palms	6.9 Ma	Habitat isolation, assortative mating	—	274 AFLPs	0.31	1.5%	Savolainen et al. (2006)
<i>Litorina saxatilis</i> “crab” and “wave”	Marine snail	0.009 Ma	Habitat isolation, assortative mating	6 allopatric, 3 regions (N = 32)	614 AFLPs	0–0.027 <sup>a</sup>	1.8–8.3%	Butlin et al. (2014), also Wilding et al. (2001), Grahame et al. (2006), Wood et al. (2008), and Galindo et al. (2010) Harr (2006)
<i>Mus musculus domesticus</i> and <i>M. m. musculus</i>	House mouse	0.5 Ma	Assortative mating, hybrid sterility	2 allopatric (N = 15)	10 265 SNPs (whole genome)	—	—	—
<i>Neochlamis bebbianae</i> “willow” and “maple”	Leaf beetle	—	Habitat isolation, assortative mating	5 allopatric (N = 165)	447 AFLPs	0.0363–0.1060 <sup>a</sup>	4.0–8.1%	Egan et al. (2008)
<i>Populus alba</i> and <i>Populus tremula</i>	Poplar	—	Assortative mating, postzygotic isolation	2 allopatric (N = 14)	38 525 SNPs (RADseq)	0.634	—	Störling et al. (2012)
<i>Quercus robur</i> and <i>Quercus petraea</i>	Oak	—	Habitat isolation	14–20 allopatric (N = 50–1190)	389 markers (isozymes, AFLPs, SCARs, microsatellites, SNPs)	0.0357	12%	Scotti-Saintagne et al. (2004)
<i>Rhagoletis pomonella</i> “apple” and “hawthorne”	Apple maggot	>0.001 Ma	Habitat isolation, assortative mating	5 allopatric (N = 508) micro satel- lites/1419 allopatric	39 microsatel- lites/ allozymes	0.0141–0.0066 <sup>a</sup>	7.7%	Michel et al. (2010), also Schwarz et al. (2009)
<i>Silene latifolia</i> and <i>Silene dioica</i>	Campion flower	—	Habitat isolation, assortative mating	6 allopatric (N = 180)	305 AFLPs	0.39–0.58 <sup>a</sup>	9.6%	Minder and Widmer (2008)
<i>Timema cristinae</i> “ <i>Ceanothus</i> ” and “ <i>Adenostoma</i> ”	Walking-stick insect	—	Habitat isolation, assortative mating	8 allopatric (N = 161)	86 130 SNPs (RADseq)	0.111	17.6%	Nosil, Parchman, et al. (2012), also Nosil et al. (2008)
<i>Zairaphera diniana</i> “larch” and “pine”	Larch budmoth	—	Habitat isolation, assortative mating	5 allopatric (N = 92)	1291 AFLPs	0.216	17.7%	Emelianov et al. (2004)

For each pair, the table not only summarizes data from the most recent study but also includes references to earlier genome scans. Only studies that estimated divergence for >20 markers are included. The columns provide information on the estimated time since divergence (Time), known barriers to gene exchange (Barriers), the number of populations sampled and total number of individuals genotyped (Populations/sample size), the number/type of markers and their source (Markers/library type), the average  $F_{ST}$  between the taxon pairs ( $F_{ST}$ ), and the number of outlier loci identified in each study (Outliers). Ma = million of years ago; N = sample size; AFLP = amplified fragment length polymorphism; EST = expressed sequence tag; RADseq = restriction site-associated DNA sequencing; RNAseq = transcriptome sequencing; SCAR = sequence characterized amplified region.

<sup>a</sup> The range of mean  $F_{ST}$  observed in comparisons of multiple species or population pairs.

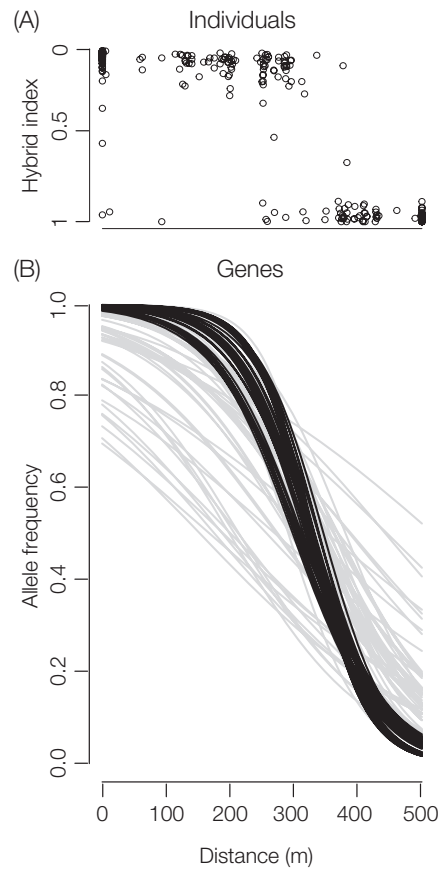
<sup>b</sup> Outliers were estimated between “benthic” and “limnetic” population pairs.

significantly more than expected; for other loci, introgression is much less than expected. Figure 2 shows an example of data from a field cricket hybrid zone, in which SNPs are chosen because they exhibit major allele frequency differences between allopatric populations that reveal variable patterns of introgression (Larson et al. 2013; Larson et al. 2014). A subset of SNPs show limited introgression in 2 very different regions of the hybrid zone and at very different spatial scales. SNPs that show consistent patterns of restricted introgression across multiple transects or contacts may mark genome regions that are components of a “universal” species boundary. Indeed, differential introgression is characteristic of all hybrid zones for which multiple markers have been studied (Table 2). Regions/alleles that introgress more than expected may be examples of adaptive introgression; regions with restricted introgression may be associated with divergent directional selection, hybrid unfitness, and/or positive assortment, that is, they may harbor genes that determine speciation phenotypes. Recent comparisons of whole-genome sequences from humans and Neanderthals have revealed evidence of hybridization and differential introgression between the 2 lineages. These studies document both reduced introgression on the X chromosome (perhaps associated with the presence of male sterility genes on the X; Sankararaman et al. 2014) and signatures of adaptive introgression for genes that determine skin phenotypes (Vernot and Akey 2014).

## The Species Boundary as a Continuum

Are species discrete entities and what is their relationship to varieties, races, and subspecies? In confronting this question, Mallet et al. (2007) and Mallet (2008a, 2008b) have repeatedly stated that Darwin (1859) got it right and that Mayr (1942, 1963) got it wrong. The essence of the argument is that Darwin emphasized continuity between varieties and species, whereas Mayr emphasized that species are real and discrete entities. Without wading into the murky waters of interpreting exactly what each of these prominent evolutionary biologists had to say, it is evident that there is truth in both points of view.

Geographic populations of the “same” species can be distinct in many ways, and such distinct populations are often recognized as races, strains, subspecies, or semispecies. A classic example of varying amounts of divergence between allopatric populations is the *Drosophila willistoni* group in South America, a group studied closely by Dobzhansky and his students (e.g., Ayala et al. 1974). Using data from 36 allozyme loci, Ayala et al. (1974) demonstrated increasing genetic differentiation (increasing proportion of loci showing significant allele frequency differences) in comparisons of conspecific geographic populations, subspecies, semispecies, sibling species, and morphologically distinct species. For these flies, and perhaps for many examples of allopatric divergence, the species boundary would seem to increase with time since divergence. An important consequence is that hybrid zone interactions can reflect a continuum of times



**Figure 2.** Differential introgression across a hybrid zone between the field crickets *Gryllus firmus* and *Gryllus pennsylvanicus*. The hybrid zone was sampled along a 500-m transect that spans the boundary between habitat patches (sand and loam soils). (A) The hybrid index (HI, estimated from 110 diagnostic SNPs) for each cricket ( $N = 260$ ) plotted against the distance along the transect. There is an abrupt transition from *G. pennsylvanicus*-like (left, HI = 0) to *G. firmus*-like (right, HI = 1) crickets at approximately 320 m. (B) The change in *G. pennsylvanicus* allele frequencies for 110 markers along the transect. Many loci have gradual changes in allele frequencies, whereas others have relatively abrupt changes that coincide with the transition between species seen in panel (A). These loci have restricted introgression relative to the other loci and represent regions of the genome that define the species boundary.

and stages of taxon divergence. That is, secondary contact following geographic isolation can occur at varying times subsequent to the vicariance or dispersal event that led to isolation.

More problematic is whether sympatric populations also reveal the same continuum. If divergence with gene flow occurs easily/regularly, then the species boundary will grow in situ, and again, we might expect a continuum. Some recent studies address this issue by comparing patterns of differentiation between ecotypes with patterns of differentiation



**Table 2** Studies of introgression across hybrid zones

Taxa compared	Organism	Time	Barriers	Populations (sample size)	Struct.	Markers (library type)	Method	Cline width	Diff	Asym	References
<i>Cottus perifretum</i> and <i>Cottus rheneanus</i>	Sculpin	1–2 Ma	Possible hybrid inviability	3 allo (N = 136), 2 sym (N = 344)	Clinal	858 micros	Genomic	NA	Y	N	Nolte et al. (2009), also Nolte et al. (2006)
<i>Gryllus firmus</i> and <i>Gryllus pennsylvanicus</i>	Field cricket	0.2 Ma	Habitat isolation, assortative matings, postmating prezygotic	6 allo (N = 71), 2 sym (N = 561)	Mosaic	168 SNPs (RNAseq)	Genomic/geographic	162–861 m	Y	Y	Larson, White, et al. (2013), also Larson, Andrés, et al. (2013)
<i>Helianthus annuus</i> and <i>Helianthus petiolaris</i>	Sunflower	1.8 Ma	Habitat isolation, hybrid sterility	6 allo (N = 69), 4 sym (N = 228)	Mosaic	110 RAPDs (linkage map)	Genomic	NA	Y	N	Buerkle and Rieseberg (2001), Gompert and Buerkle (2009), also Rieseberg et al. (1999)
<i>Lycæides idas</i> and <i>Lycæides melissa</i>	Butterfly	2.4 Ma	Habitat isolation, possible premating barriers	2 allo (N = 192), 1 sym (N = 186)	Patchy	119 677 SNPs (RADseq)	Genomic	NA	Y	Y	Gompert et al. (2012)
<i>Manacus candei</i> and <i>Manacus vitellinus</i>	Manakin	—	Assortative mating	2 allo (N = 100), 1 sym (N = 104)	Clinal	119 677 SNPs (RADseq)	Genomic	NA	Y	N	Parchman et al. (2013)
<i>Mus musculus domesticus</i> and <i>M. m. musculus</i>	Mouse	0.5 Ma	Assortative mating, hybrid sterility	2 allo (N = 14), 2 sym (N = 679)	Clinal	59 100 SNPs (whole genome)	Genomic/geographic	6.4–341.8 km <sup>a</sup>	Y	Y	Janousek et al. (2012), also Payseur et al. (2004), Teeter et al. (2008, 2010), and Macholán et al. (2011)
<i>Oryctolagus c. cuniculus</i> and <i>Oryctolagus c. algirus</i>	Rabbit	1.8 Ma	Unknown	Transsect (N = 1078)	Clinal	1401 SNPs (whole genome)	Geographic	10–545 km	Y	N	Carneiro et al. (2013), also Carneiro et al. (2010)
<i>Picea sitchensis</i> and <i>Picea glauca</i>	Spruce	—	Habitat isolation	2 allo (N = 66), 29 sym (N = 721)	Clinal	22 SNPs (can-didate genes)	Genomic	NA	Y	Y	Hamilton et al. (2013a, 2013b)
<i>Populus alba</i> and <i>Populus tremula</i>	Poplar	—	Habitat isolation, possible post-mating barriers	6 allo (N = 248), 3 sym (N = 436)	Mosaic	268 micros (whole genome)	Genomic	NA	Y	Y	Lindtke et al. (2012), also Lexer et al. (2007, 2010)
<i>Timema cristinae</i> “ <i>Ceanothus</i> ” and “ <i>Adenostoma</i> ”	Walking-stick insect	—	Habitat isolation, assortative mating	2 allo (N = 42), 2 para (N = 84)	Parapatric	38 304 SNPs (RADseq)	Genomic	NA	Y	N	Nosil, Gompert, et al. (2012)

For each hybrid zone, data from the most recent study are summarized, but references to earlier studies are included. Only hybrid zones for which >20 markers were analyzed are included. For each pair of hybridizing taxa, the table includes estimated divergence time (Time), documented barriers to gene exchange (Barriers), the number of populations sampled and total number of individuals genotyped (Populations), the hybrid zone structure (Struct.), the number/type of markers and their source (Markers/library type), the method of estimating introgression (Method), the estimated geographic cline width (cline width), whether the studies found evidence of differential introgression (Diff) or asymmetric introgression (Asym) and references. N = sample size; allo = allopatric populations; sym = sympatric populations; para = parapatric populations; micros = microsatellite loci; RADseq = restriction site-associated DNA sequencing; RAPD = random amplified polymorphic DNA.

<sup>a</sup> Cline width estimated from Teeter et al. (2008).

between species (e.g., [Andrew and Rieseberg 2013](#)). Because Mayr did not believe that sympatric speciation occurred (except in a few cases, e.g., allopolyploid hybrid speciation in plants), he emphasized discontinuity. In fact, species histories are generally more complex than simple allopatric or sympatric scenarios might have us believe, and many taxa have probably experienced alternating periods of isolation and gene flow. In the end, it seems that one does not have to take sides in the Darwin versus Mayr debate to recognize the species boundary as a continuum.

## Hybrid Speciation, Adaptive Introgression, and the Origin of Novel Traits

Hybridization may contribute directly to the origin of species, either as a result of reinforcement or hybrid speciation ([Servedio and Noor 2003](#); [Mallet 2007](#); [Abbott et al. 2010, 2013](#)). Some proponents of this view, like many of their colleagues, invoke the specter of [Mayr \(1942\)](#) and suggest that hybridization has traditionally been viewed as an “evolutionary dead end” ([Seehausen 2013](#)), or together with gene flow, as “mainly destructive forces with little evolutionary consequence” ([Saetre 2013](#)). Homoploid hybrid speciation involves the formation of novel genetic combinations and novel adaptations that allow persistence of the hybrid lineage, often in an environment distinct from that of either parent. Recognized as a common phenomenon in plants ([Arnold 1997](#); [Abbott et al. 2010](#)), homoploid hybrid speciation has more recently gained support as a speciation mechanism in animals ([Gompert et al. 2006](#); [Mallet 2007](#)). However, this potentially “constructive” role for hybridization remains controversial and many think that homoploid hybrid speciation will not turn out to be an important mode of speciation in animals ([Barton 2013](#); [Servedio et al. 2013](#); [Schumer et al. 2014](#)).

One of the contentious issues is the relative contribution of hybridization (vs. mutation) as a source of novel alleles or genotypes. Hybridization allows introgression of combinations of alleles that have already been “tested” by natural selection. Moreover, because of the greater genetic differences between (as opposed to within) hybridizing taxa, one outcome of hybridization may be the appearance of transgressive phenotypes (extreme phenotypes not seen in either of the parents), which is a source of evolutionary novelty. But it can also be argued that in a set of populations subdivided by hybrid zones, novel adaptations will appear no faster than if the entire set of populations was panmictic ([Barton 2013](#)). Similarly, hybridization tends to make 2 populations more similar (not less) and therefore must (at some level) oppose divergence of the hybridizing lineages ([Servedio et al. 2013](#)). It is possible that adaptive introgression of traits from species A into species B might lead to splitting of B into B and B', that is, the introgression of traits from A may render some individuals of B sufficiently different from others that they are now effectively 2 species. This appears to be the case in *Heliconius* butterflies, where alleles at loci encoding wing

color patterns have introgressed ([Pardo-Diaz et al. 2012](#)). Numerous examples of adaptive introgression have been reported, but few result in speciation events.

## Human-Mediated Secondary Contact

Semipermeable species boundaries have important implications for human-mediated secondary contact. Such contact may occur as a consequence of environmental disturbance, accidental introductions, or intentional introductions of wild populations, crop plants, or domestic animals. Thus, introduced species may overlap with and potentially interbreed with congeners, and in some of these cases, there is evidence for differential introgression ([Abbott et al. 2003](#); [McDonald et al. 2008](#); [Feulner et al. 2013](#); [Goedbloed et al. 2013](#); [Hohenlohe et al. 2013](#)). Gene flow may carry alleles in both directions (from introduced into native and vice versa), and the consequences of gene flow may be problematic. For example, transgenes or other alleles from crop plants can make their way into populations of wild relatives. These alleles may increase the fitness of the wild plants, and thus, natural selection will drive introgression ([Ellstrand 2003](#); [Snow et al. 2010](#); [Snow 2012](#)). Similarly, hybridization between domesticated sheep and their wild relatives has resulted in changes in coat color and pattern in the wild sheep, changes that appear to be adaptive ([Feulner et al. 2013](#)).

But “adaptive” changes in wild populations may not be desirable; alleles that confer resistance to pesticides or herbicides may endow insects or plants with properties that allow them to flourish but that we view with concern (e.g., a weedy plant becomes resistant to herbivory; [Yang et al. 2011](#)). The probability that such transfers will occur depends not only on the selective advantage/disadvantage conferred by a particular allele but also on the genomic location of the gene and its linkage relationship to other genes. In situations where human-mediated secondary contact allows for hybridization and introgression between species that previously were not connected by gene flow, genome scans define the genomic context in which potentially invasive alleles are embedded and thereby provide information about the likelihood of introgression ([Hohenlohe et al. 2013](#)).

## Conclusions

Patterns of differentiation between recently diverged taxa and patterns of variation in hybrid zones provide important insights into the genetic architecture of species boundaries. For the past 40 years, evolutionary biologists have been using molecular markers to characterize differentiation between species and races and to define allele and genotype frequencies across natural hybrid zones. In the beginning, markers were few and reference genomes unimagined. Today, markers are virtually unlimited in number and reference genomes relatively easy to obtain. More and better genetic data can be obtained in a single Illumina Hi-Seq run than could be obtained over many years of using RFLPs or other indirect

methods for assaying DNA sequence variation. These data now allow genome-wide patterns of divergence or differential introgression to be described in remarkable detail, although in few cases are convincing explanations for these patterns available.

In contrast to the major advances in data generation and analysis, the questions being asked by evolutionary biologists have remained much the same. Recent data reinforce conclusions based on many fewer loci: species boundaries are semipermeable, with permeability varying as a function of genome region. Thus, hybridizing taxa often remain distinct for only part of the genome. The proportion of the genome that is resistant to introgression varies among taxa and, in some cases, patterns of introgression appear to be different when data are available for multiple transects across the “same” hybrid zone (e.g., [Teeter et al. 2010](#)). This suggests that the species boundary may vary geographically, perhaps a result of local adaptation in heterogeneous environments. Genome regions that consistently show reduced introgression between pairs of hybridizing taxa likely harbor genes that contribute to barriers that are independent of environmental variation. Working with sunflowers, [Rieseberg et al. \(1999\)](#) clearly documented such a pattern, and more recent work on a field cricket hybrid zone has identified a set of markers that exhibit reduced introgression in 2 distinct regions of the hybrid zone ([Larson et al. 2014](#)).

Genomic divergence and differential introgression are likely taxon specific, but some consistent patterns have begun to emerge. The massive amounts of data that are now being produced in a wide variety of natural systems promise that we may soon have a clearer picture of the details of species boundaries. Comparisons of diverging lineages provide static views of patterns of differentiation across the genome, but with more data, we ultimately will be able to define the dynamics of species boundaries, how boundaries become less (or more) permeable over time.

## Funding

National Science Foundation and United States Department of Agriculture (to R.G.H.). Over many years, these sources have funded research on a field cricket hybrid zone (National Science Foundation) and gene exchange between pheromone strains of the European Corn Borer (United States Department of Agriculture and National Science Foundation). These hybridizing insects have provided the context in which our thinking about the nature of species boundaries has evolved.

## Acknowledgments

Many members (past and present) of the Harrison lab have helped us to refine our thinking about hybrid zones and species boundaries. M. Hahn and 2 anonymous reviewers provided insightful comments that have substantially improved the manuscript. We thank K. Shaw and others at the American Genetic Association for hosting the symposium on speciation from which this paper has emerged.

## References

- Abbott RJ, Albach D, Ansell S, Arntzen JW, Baird SJ, Bierne N, Boughman JW, Brelsford A, Buerkle AC, Buggs R, et al. 2013. Hybridization and speciation. *J Evol Biol.* 26:229–246.
- Abbott RJ, Hegarty MJ, Hiscock SJ, Brennan AC. 2010. Homoploid hybrid speciation in action. *Taxon.* 59:1375–1386.
- Abbott RJ, James JK, Milne RI, Gillies ACM. 2003. Plant introductions, hybridization and gene flow. *Philos Trans R Soc B.* 358:1123–1132.
- Anderson E. 1949. *Introgressive hybridization*. New York: Wiley & Sons.
- Anderson E, Hubricht L. 1938. Hybridization in *Tradescantia*. III. The evidence for introgressive hybridization. *Am J Bot.* 25:396–402.
- Andrés JA, Larson EL, Bogdanowicz S, Harrison RG. 2013. Patterns of transcriptome divergence in the male accessory gland of two closely related species of field crickets. *Genetics.* 193:501–513.
- Andrés JA, Maroja LS, Harrison RG. 2008. Searching for candidate speciation genes using a proteomic approach: seminal proteins in field crickets. *Proc R Soc B.* 275:1975–1983.
- Andrew RL, Rieseberg LH. 2013. Divergence is focused on few genomic regions early in speciation: incipient speciation of sunflower ecotypes. *Evolution.* 67:2468–2482.
- Arnold M. 1997. *Natural hybridization and evolution*. New York: Oxford University Press.
- Ayala F, Tracey M, Hedgecock D. 1974. Genetic differentiation during the speciation process in *Drosophila*. *Evolution.* 28:576–592.
- Barton NH, Gale KH. 1993. Genetic analysis of hybrid zones. In: Harrison RG, editor. *Hybrid zones and the evolutionary process*. New York: Oxford University Press. p. 13–45.
- Barton NH, Hewitt GM. 1981. Hybrid zones and speciation. In: Atchley WR, Woodruff DS, editors. *Evolution and speciation*. Cambridge (UK): Cambridge University Press. p. 109–145.
- Barton NH, Hewitt GM. 1985. Analysis of hybrid zones. *Annu Rev Ecol Syst.* 16:113–148.
- Barton N, Jones JS. 1983. Mitochondrial DNA: new clues about evolution. *Nature.* 306:317–318.
- Barton NH. 1979. Gene flow past a cline. *Heredity.* 43:333–339.
- Barton NH. 2013. Does hybridization influence speciation? *J Evol Biol.* 26:267–269.
- Bazykin A. 1969. Hypothetical mechanism of speciation. *Evolution.* 23:685–687.
- Beaumont MA, Balding DJ. 2004. Identifying adaptive genetic divergence among populations from genome scans. *Mol Ecol.* 13:969–980.
- Beltran M, Jiggins CD, Bull V, Linares M, Mallet J, McMillan OW, Bermingham E. 2002. Phylogenetic discordance at the species boundary: comparative gene genealogies among rapidly radiating *Heliconius* butterflies. *Mol Biol Evol.* 19:2176–2190.
- Bouck A, Peeler R, Arnold ML, Wessler SR. 2005. Genetic mapping of species boundaries in Louisiana irises using IRRE retrotransposon display markers. *Genetics.* 171:1289–1303.
- Buerkle AC, Rieseberg LH. 2001. Low intraspecific variation for genomic isolation between hybridizing sunflower species. *Evolution.* 55:684–691.
- Butlin RK, Saura M, Charrier G, Jackson B, André C, Caballero A, Coyne JA, Galindo J, Grahame JW, Hollander J, et al. 2014. Parallel evolution of local adaptation and reproductive isolation in the face of gene flow. *Evolution.* doi:10.1111/evo.12329.
- Campbell D, Bernatchez L. 2004. Generic scan using AFLP markers as a means to assess the role of directional selection in the divergence of sympatric whitefish ecotypes. *Mol Biol Evol.* 21:945–956.
- Carneiro M, Baird SJ, Afonso S, Ramirez E, Tarroso P, Teotónio H, Villafuerte R, Nachman MW, Ferrand N. 2013. Steep clines within a highly

- permeable genome across a hybrid zone between two subspecies of the European rabbit. *Mol Ecol*. 22:2511–2525.
- Carneiro M, Blanco-Aguilar JA, Villafuerte R, Ferrand N, Nachman MW. 2010. Speciation in the European rabbit (*Oryctolagus cuniculus*): islands of differentiation on the X chromosome and autosomes. *Evolution*. 64:3443–3460.
- Charlesworth B. 1998. Measures of divergence between populations and the effect of forces that reduce variability. *Mol Biol Evol*. 15:538–543.
- Coyne JA, Orr H. 2004. *Speciation*. Sunderland (MA): Sinauer Associates, Inc.
- Cruikshank TE, Hahn MW. Forthcoming 2014. Reanalysis suggests that genomic islands of speciation are due to reduced diversity, not reduced gene flow. *Mol Ecol*.
- Cui R, Schumer M, Kruesi K, Walter R, Andolfatto P, Rosenthal GG. 2013. Phylogenomics reveals extensive reticulate evolution in *Xiphophorus* fishes. *Evolution*. 67:2166–2179.
- Darwin C. 1859. *On the origin of species by natural selection*. London: Murray.
- Deagle BE, Jones FC, Chan YF, Absher DM, Kingsley DM, Reimchen TE. 2012. Population genomics of parallel phenotypic evolution in stickleback across stream-lake ecological transitions. *Proc R Soc B*. 279:1277–1286.
- Dopman E, Perez L, Bogdanowicz S, Harrison RG. 2005. Consequences of reproductive barriers for genealogical discordance in the European corn borer. *Proc Natl Acad Sci USA*. 102:14706–14711.
- Egan SP, Nosil P, Funk DJ. 2008. Selection and genomic differentiation during ecological speciation: isolating the contributions of host association via a comparative genome scan of *Neochlamisus bebbianae* leaf beetles. *Evolution*. 62:1162–1181.
- Ellegren H, Smeds L, Burri R, Olason PI, Backström N, Kawakami T, Künstner A, Mäkinen H, Nadachowska-Brzyska K, Qvarnström A, et al. 2012. The genomic landscape of species divergence in *Ficedula* flycatchers. *Nature*. 491:756–760.
- Ellstrand NC. 2003. Current knowledge of gene flow in plants: implications for transgene flow. *Philos Trans R Soc B*. 358:1163–1170.
- Emelianov I, Marec F, Mallet J. 2004. Genomic evidence for divergence with gene flow in host races of the larch budmoth. *Proc R Soc B*. 271:97–105.
- Feder JL, Egan SP, Nosil P. 2012. The genomics of speciation-with-gene-flow. *Trends Genet*. 28:342–350.
- Feder JL, Nosil P. 2010. The efficacy of divergence hitchhiking in generating genomic islands during ecological speciation. *Evolution*. 64:1729–1747.
- Ferris SD, Sage RD, Huang CM, Nielsen JT, Ritte U, Wilson AC. 1983. Flow of mitochondrial DNA across a species boundary. *Proc Natl Acad Sci USA*. 80:2290–2294.
- Feulner PGD, Gratten J, Kijas JW, Visscher PM, Pemberton JM, Slate J. 2013. Introgression and the fate of domesticated genes in a wild mammal population. *Mol Ecol*. 22:4210–4221.
- Fitzpatrick BM. 2013. Alternative forms for genomic clines. *Ecol Evol*. doi:10.1002/ece3.609.
- Flaxman SM, Feder JL, Nosil P. 2012. Spatially explicit models of divergence and genome hitchhiking. *J Evol Biol*. 25:2633–2650.
- Gagnaire PA, Pavey SA, Normandeau E, Bernatchez L. 2013. The genetic architecture of reproductive isolation during speciation-with-gene-flow in lake whitefish species pairs assessed by RAD sequencing. *Evolution*. 67:2483–2497.
- Galindo J, Grahame JW, Butlin RK. 2010. An EST-based genome scan using 454 sequencing in the marine snail *Littorina saxatilis*. *J Evol Biol*. 23:2004–2016.
- Garrigan D, Kingan SB, Geneva AJ, Andolfatto P, Clark AG, Thornton KR, Presgraves DC. 2012. Genome sequencing reveals complex speciation in the *Drosophila simulans* clade. *Genome Res*. 22:1499–1511.
- Goedbloed DJ, Megens HJ, van Hooft P, Herrero-Medrano JM, Lutz W, Alexandris P, Crooijmans RPMA, Groenen M, Wieren SE, Ydenberg RC, et al. 2013. Genome-wide single nucleotide polymorphism analysis reveals recent genetic introgression from domestic pigs into Northwest European wild boar populations. *Mol Ecol*. 22:856–866.
- Gompert Z, Buerkle AC. 2009. A powerful regression-based method for admixture mapping of isolation across the genome of hybrids. *Mol Ecol*. 18:1207–1224.
- Gompert Z, Buerkle AC. 2011. Bayesian estimation of genomic clines. *Mol Ecol*. 20:2111–2127.
- Gompert Z, Buerkle AC. 2012. beg: software for Bayesian estimation of genomic clines. *Mol Ecol Res*. 12:1168–1176.
- Gompert Z, Fordyce JA, Forister ML, Shapiro AM, Nice CC. 2006. Homoploid hybrid speciation in an extreme habitat. *Science*. 314:1923–1925.
- Gompert Z, Lucas LK, Nice CC, Fordyce JA, Forister ML, Buerkle AC. 2012. Genomic regions with a history of divergent selection affect fitness of hybrids between two butterfly species. *Evolution*. 66:2167–2181.
- Grahame JW, Wilding C, Butlin RK. 2006. Adaptation to a steep environmental gradient and an associated barrier to gene exchange in *Littorina saxatilis*. *Evolution*. 60:268–278.
- Grant PR, Grant BR. 2010. Conspecific versus heterospecific gene exchange between populations of Darwin's finches. *Philos Trans R Soc B*. 365:1065–1076.
- Hamilton JA, Lexer C, Aitken SN. 2013a. Differential introgression reveals candidate genes for selection across a spruce (*Picea sitchensis* × *P. glauca*) hybrid zone. *New Phytol*. 197:927–938.
- Hamilton JA, Lexer C, Aitken SN. 2013b. Genomic and phenotypic architecture of a spruce hybrid zone (*Picea sitchensis* × *P. glauca*). *Mol Ecol*. 22:827–841.
- Harr B. 2006. Genomic islands of differentiation between house mouse subspecies. *Genome Res*. 16:730–737.
- Harrison RG. 1989. Animal mitochondrial DNA as a genetic marker in population and evolutionary biology. *Trends Ecol Evol*. 4:6–11.
- Harrison RG. 1990. Hybrid zones: windows on evolutionary process. In: Futuyma D, Antonovics J, editors. *Oxford surveys in evolutionary biology*. Vol. 7. New York: Oxford University Press. p. 69–128.
- Harrison RG, editor. 1993. *Hybrid zones and the evolutionary process*. New York: Oxford University Press.
- Harrison RG. 1998. Linking evolutionary pattern and process: the relevance of species concepts for the study of speciation. In: Howard DJ, Berlocher SH, editors. *Endless forms species and speciation*. New York: Oxford University Press. p. 19–31.
- Harrison RG. 2012. The language of speciation. *Evolution*. 66:3643–3657.
- Harrison RG, Rand DM, Wheeler WC. 1987. Mitochondrial DNA variation in field crickets across a narrow hybrid zone. *Mol Biol Evol*. 4:144–158.
- Hebert FO, Renaut S, Bernatchez L. 2013. Targeted sequence capture and resequencing implies a predominant role of regulatory regions in the divergence of a sympatric lake whitefish species pair (*Coregonus clupeaformis*). *Mol Ecol*. 22:4896–4914.
- Hedrick PW. 2013. Adaptive introgression in animals: examples and comparison to new mutation and standing variation as sources of adaptive variation. *Mol Ecol*. 22:4606–4618.
- Heiser CB. 1973. Introgression re-examined. *Bot Rev*. 39:347–366.
- Hohenlohe P, Bassham S, Currey M, Cresko W. 2012. Extensive linkage disequilibrium and parallel adaptive divergence across threespine stickleback genomes. *Philos Trans R Soc B*. 367:395–408.
- Hohenlohe P, Bassham S, Etter PD, Stiffler N, Johnson EA, Cresko W. 2010. Population genomics of parallel adaptation in threespine stickleback using sequenced RAD tags. *PLoS Genet*. 6:e1000862.
- Hohenlohe P, Day MD, Amish SJ, Miller MR, Kamps-Hughes N, Boyer MC, Muhlfeld CC, Allendorf FW, Johnson EA, Luikart G. 2013. Genomic

- patterns of introgression in rainbow and westslope cutthroat trout illuminated by overlapping paired-end RAD sequencing. *Mol Ecol.* 22:3002–3013.
- Hudson RR, Coyne JA. 2002. Mathematical consequences of the geological species concept. *Evolution.* 56:1557–1565.
- Janousek V, Wang L, Luzynski K, Dufková P, Vyskocilová MM, Nachman MW, Munclinger P, Macholán M, Piálek J, Tucker P. 2012. Genome-wide architecture of reproductive isolation in a naturally occurring hybrid zone between *Mus musculus musculus* and *M. m. domesticus*. *Mol Ecol.* 21:3032–3047.
- Jaquière J, Stoeckel S, Nouhaud P, Mieuze L, Mahéo F, Legeai F, Bernard N, Bonvoisin A, Vitalis R, Simon JC. 2012. Genome scans reveal candidate regions involved in the adaptation to host plant in the pea aphid complex. *Mol Ecol.* 21:5251–5264.
- Jones FC, Chan YF, Schmutz J, Grimwood J, Brady SD, Southwick AM, Absher DM, Myers RM, Reimchen TE, Deagle BE, et al. 2012. A genome-wide SNP genotyping array reveals patterns of global and repeated species-divergence in sticklebacks. *Curr Biol.* 22:83–90.
- Jones FC, Grabherr MG, Chan YF, Russell P, Mauceci E, Johnson J, Swofford R, Pirun M, Zody MC, White S, et al. 2012. The genomic basis of adaptive evolution in threespine sticklebacks. *Nature.* 484:55–61.
- Kane NC, King MG, Barker MS, Raduski A, Karrenberg S, Yatabe Y, Knapp SJ, Rieseberg LH. 2009. Comparative genomic and population genetic analyses indicate highly porous genomes and high levels of gene flow between divergent *Helianthus* species. *Evolution.* 63:2061–2075.
- Keller I, Wagner CE, Greuter L, Mwaiko S, Selz OM, Sivasundar A, Wittwer S, Seehausen O. 2013. Population genomic signatures of divergent adaptation, gene flow and hybrid speciation in the rapid radiation of Lake Victoria cichlid fishes. *Mol Ecol.* 22:2848–2863.
- Key K. 1968. The concept of stasipatric speciation. *Syst Biol.* 17:14–22.
- Larson EL, Andrés JA, Bogdanowicz S, Harrison RG. 2013. Differential introgression in a mosaic hybrid zone reveals candidate barrier genes. *Evolution.* 67:3653–3651.
- Larson EL, White TA, Ross C, Harrison RG. 2014. Gene flow and the maintenance of species boundaries. *Mol Ecol.* 23:1668–1678.
- Lawniczak M, Emrich SJ, Holloway AK, Regier AP, Olson M, White M, Redmond S, Fulton L, Appelbaum E, Farmer C, et al. 2010. Widespread divergence between incipient *Anopheles gambiae* species revealed by whole genome sequences. *Science.* 330:512–514.
- Lemmon EM, Lemmon AR, Collins JT, Lee-Yaw JA, Cannatella DC. 2007. Phylogeny-based delimitation of species boundaries and contact zones in the trilling chorus frogs (*Pseudacris*). *Mol Phylogenet Evol.* 44:1068–1082.
- Lexer C, Joseph J, Barbara T, van Loo M, Heinze B, Bartha D, Castiglione S, Fay MF, Buerkle AC. 2010. Genomic admixture analysis in European *Populus* spp. reveals unexpected patterns of reproductive isolation and mating. *Genetics.* 186:699–712.
- Lexer C, Joseph J, Buerkle AC, Heinze B, Fay MF. 2007. Admixture in European *Populus* hybrid zones makes feasible the mapping of loci that contribute to reproductive isolation and trait differences. *Heredity.* 98:74–84.
- Lindtke D, Buerkle AC, Barará T, Heinze B, Castiglione S, Bartha D, Lexer C. 2012. Recombinant hybrids retain heterozygosity at many loci: new insights into the genomics of reproductive isolation in *Populus*. *Mol Ecol.* 21:5042–5058.
- Machado CA, Hey J. 2003. The causes of phylogenetic conflict in a classic *Drosophila* species group. *Proc R Soc B.* 270:1193–1202.
- Macholán M, Baird SJ, Dufková P, Munclinger P, Bímová B, Piálek J. 2011. Assessing multilocus introgression patterns: a case study on the mouse X chromosome in central Europe. *Evolution.* 65:1428–1446.
- Mallet J. 2007. Hybrid speciation. *Nature.* 446:279–283.
- Mallet J. 2008a. Mayr's view of Darwin: was Darwin wrong about speciation? *Biol J Linn Soc.* 95:3–16.
- Mallet J. 2008b. Hybridization, ecological races and the nature of species: empirical evidence for the ease of speciation. *Philos Trans R Soc B.* 363:2971–2986.
- Mallet J, Beltrán M, Neukirchen W, Linares M. 2007. Natural hybridization in heliconiine butterflies: the species boundary as a continuum. *BMC Evol Biol.* 7:1–16.
- Mayr E. 1942. Systematics and the origin of species, from the viewpoint of a zoologist. Cambridge (MA): Harvard University Press.
- Mayr E. 1963. Animal species and evolution. Cambridge (MA): Harvard University Press.
- Mayr E. 1982. The growth of biological thought: diversity, evolution, and inheritance. Cambridge (MA): Belknap Press.
- McDonald DB, Parchman TL, Bower MR, Hubert WA, Rahel FJ. 2008. An introduced and a native vertebrate hybridize to form a genetic bridge to a second native species. *Proc Natl Acad Sci USA.* 105:10837–10842.
- Michel AP, Sim S, Powell THQ, Taylor MS, Nosil P, Feder JL. 2010. Widespread genomic divergence during sympatric speciation. *Proc Natl Acad Sci USA.* 107:9724–9729.
- Minder AM, Widmer A. 2008. A population genomic analysis of species boundaries: neutral processes, adaptive divergence and introgression between two hybridizing plant species. *Mol Ecol.* 17:1552–1563.
- Nachman MW, Payseur BA. 2012. Recombination rate variation and speciation: theoretical predictions and empirical results from rabbits and mice. *Philos Trans R Soc B.* 367:409–421.
- Nadeau NJ, Martin SH, Kozak KM, Salazar C, Dasmahapatra KK, Davey JW, Baxter SW, Blaxter ML, Mallet J, Jiggins CD. 2013. Genome-wide patterns of divergence and gene flow across a butterfly radiation. *Mol Ecol.* 22:814–826.
- Nadeau NJ, Whibley A, Jones RT, Davey JW, Dasmahapatra KK, Baxter SW, Quail MA, Joron M, French-Constant RH, Blaxter ML, et al. 2012. Genomic islands of divergence in hybridizing *Heliconius* butterflies identified by large-scale targeted sequencing. *Philos Trans R Soc B.* 367:343–353.
- Navarro A, Barton NH. 2003. Chromosomal speciation and molecular divergence: accelerated evolution in rearranged chromosomes. *Science.* 300:321–324.
- Neafsey DE, Lawniczak M, Park DJ, Redmond S, Coulibaly MB, Traore SF, Sagnon N, Costantini C, Johnson C, Wiegand RC, et al. 2010. SNP genotyping defines complex gene-flow boundaries among African malaria vector mosquitoes. *Science.* 330:514–517.
- Nolte AW, Freyhof J, Tautz D. 2006. When invaders meet locally adapted types: rapid moulding of hybrid zones between sculpins (*Cottus*, Pisces) in the Rhine system. *Mol Ecol.* 15:1983–1993.
- Nolte AW, Gompert Z, Buerkle AC. 2009. Variable patterns of introgression in two sculpin hybrid zones suggest that genomic isolation differs among populations. *Mol Ecol.* 18:2615–2627.
- Noor MAF, Grams KL, Bertucci LA, Reiland J. 2001. Chromosomal inversions and the reproductive isolation of species. *Proc Natl Acad Sci USA.* 98:12084–12088.
- Nosil P, Egan SP, Funk DJ. 2008. Heterogeneous genomic differentiation between walking-stick ecotypes: 'isolation by adaptation' and multiple roles for divergent selection. *Evolution.* 62:316–336.
- Nosil P, Funk DJ, Ortiz-Barrientos D. 2009. Divergent selection and heterogeneous genomic divergence. *Mol Ecol.* 18:375–402.
- Nosil P, Gompert Z, Farkas TE, Comeault AA, Feder JL, Buerkle AC, Parchman TL. 2012. Genomic consequences of multiple speciation processes in a stick insect. *Proc R Soc B.* 279:5058–5065.
- Nosil P, Parchman TL, Feder JL, Gompert Z. 2012. Do highly divergent loci reside in genomic regions affecting reproductive isolation? A test using next-generation sequence data in *Timema* stick insects. *BMC Evol Biol.* 12:1–12.
- Parchman TL, Gompert Z, Braun MJ, Brumfield RT, McDonald DB, Uy JAC, Zhang G, Jarvis ED, Schlinger BA, Buerkle AC. 2013. The genomic consequences of adaptive divergence and reproductive isolation between species of manakins. *Mol Ecol.* 22:3304–3317.

- Pardo-Diaz C, Salazar C, Baxter SW, Merot C, Figueiredo-Ready W, Joron M, McMillan OW, Jiggins CD. 2012. Adaptive introgression across species boundaries in *Heliconius* butterflies. *Plos Genet.* 8:e1002752.
- Patterson N, Richter DJ, Gnerre S, Lander ES, Reich D. 2006. Genetic evidence for complex speciation of humans and chimpanzees. *Nature.* 441:1103–1108.
- Payseur BA. 2010. Using differential introgression in hybrid zones to identify genomic regions involved in speciation. *Mol Ecol Res.* 10:806–820.
- Payseur BA, Krenz JG, Nachman MW. 2004. Differential patterns of introgression across the X chromosome in a hybrid zone between two species of house mice. *Evolution.* 58:2064–2078.
- Powell JR. 1983. Interspecific cytoplasmic gene flow in the absence of nuclear gene flow: evidence from *Drosophila*. *Proc Natl Acad Sci USA.* 80:492–495.
- Prüfer K, Racimo F, Patterson N, Jay F, Sankararaman S, Sawyer S, Heinze A, Renaud G, Sudmant P, de Filippo C, et al. 2014. The complete genome sequence of a Neanderthal from the Altai Mountains. *Nature.* 505:43–49.
- Putnam AS, Scriber JM, Andolfatto P. 2007. Discordant divergence times among Z-chromosome regions between two ecologically distinct swallowtail butterfly species. *Evolution.* 61:912–927.
- Renaud S, Nolte AW, Bernatchez L. 2010. Mining transcriptome sequences towards identifying adaptive single nucleotide polymorphisms in lake whitefish species pairs (*Coregonus* spp. Salmonidae). *Mol Ecol.* 19:115–131.
- Renaud S, Nolte AW, Rogers SM, Derome N, Bernatchez L. 2011. SNP signatures of selection on standing genetic variation and their association with adaptive phenotypes along gradients of ecological speciation in lake whitefish species pairs (*Coregonus* spp.). *Mol Ecol.* 20:545–559.
- Rieseberg LH. 2001. Chromosomal rearrangements and speciation. *Trends Ecol Evol.* 16:351–358.
- Rieseberg LH, Whitton J, Gardner K. 1999. Hybrid zones and the genetic architecture of a barrier to gene flow between two sunflower species. *Genetics.* 152:713–727.
- Roe AD, Sperling FAH. 2007. Population structure and species boundary delimitation of cryptic *Doryctria* moths: an integrative approach. *Mol Ecol.* 16:3617–3633.
- Roesti M, Hendry AP, Salzburger W, Berner D. 2012. Genome divergence during evolutionary diversification as revealed in replicate lake-stream stickleback population pairs. *Mol Ecol.* 21:2852–2862.
- Saetre GP. 2013. Hybridization is important in evolution, but is speciation? *J Evol Biol.* 26:256–258.
- Sankararaman S, Mallick S, Dannemann M, Prüfer K, Kelso J, Paabo S, Patterson N, Reich D. 2014. The genomic landscape of Neanderthal ancestry in present-day humans. *Nature.* 507:354–357.
- Savolainen V, Anstett M-C, Lexer C, Hutton I, Clarkson JJ, Norup MV, Powell MP, Springate D, Salamin N, Baker WJ. 2006. Sympatric speciation in palms on an oceanic island. *Nature.* 441:210–213.
- Scascitelli M, Whitney KD, Randell RA, King M, Buerkle AC, Rieseberg LH. 2010. Genome scan of hybridizing sunflowers from Texas (*Helianthus annuus* and *H. debilis*) reveals asymmetric patterns of introgression and small islands of genomic differentiation. *Mol Ecol.* 19:521–541.
- Schumer M, Rosenthal G, Andolfatto P. 2014. How common is homoploid hybrid speciation? *Evolution.* doi:10.1111/evo.12399.
- Schwarz D, Robertson HM, Feder JL, Varala K, Hudson ME, Ragland GJ, Hahn DA, Berlocher SH. 2009. Sympatric ecological speciation meets pyrosequencing: sampling the transcriptome of the apple maggot *Rhagoletis pomonella*. *BMC Genomics.* 10:633.
- Scotti-Saintagne C, Mariette S, Porth I, Goicoechea PG, Barreneche T, Bodenes C, Burg K, Kremer A. 2004. Genome scanning for interspecific differentiation between two closely related oak species [*Quercus robur* L. and *Q. petraea* (Matt.) Liebl.]. *Genetics.* 168:1615–1626.
- Seehausen O. 2013. Conditions when hybridization might predispose populations for adaptive radiation. *J Evol Biol.* 26:279–281.
- Servedio MR, Hermansen JS, Van Doorn GS. 2013. Hybridization may rarely promote speciation. *J Evol Biol.* 26:282–285.
- Servedio MR, Noor MAF. 2003. The role of reinforcement in speciation: theory and data. *Annu Rev Ecol Evol Syst.* 34:339–364.
- Shaw KL, Mullen S. 2011. Genes versus phenotypes in the study of speciation. *Genetica.* 139:649–661.
- Smadja CM, Canbäck B, Vitalis R, Gautier M, Ferrari J, Zhou J-J, Butlin RK. 2012. Large-scale candidate gene scan reveals the role of chemoreceptor genes in host plant specialization and speciation in the pea aphid. *Evolution.* 66:2723–2738.
- Snow AA. 2012. Illegal gene flow from transgenic creeping bentgrass: the saga continues. *Mol Ecol.* 21:4663–4664.
- Snow AA, Culley TM, Campbell LG, Sweeney PM, Hegde SG, Ellstrand NC. 2010. Long-term persistence of crop alleles in weedy populations of wild radish (*Raphanus raphanistrum*). *New Phytol.* 186:537–548.
- Stöltig KN, Nipper R, Lindtke D, Caseys C, Waeber S, Castiglione S, Lexer C. 2012. Genomic scan for single nucleotide polymorphisms reveals patterns of divergence and gene flow between ecologically divergent species. *Mol Ecol.* 22:842–855.
- Strausberg RL, Scotti-Saintagne C, Scotti I, Lai Z, Rieseberg LH. 2009. Genomic patterns of adaptive divergence between chromosomally differentiated sunflower species. *Mol Biol Evol.* 26:1341–1355.
- Teeter KC, Payseur BA, Harris LW, Bakewell MA, Thibodeau LM, O'Brien JE, Krenz JG, Sans-Fuentes MA, Nachman MW, Tucker P. 2008. Genome-wide patterns of gene flow across a house mouse hybrid zone. *Genome Res.* 18:67–76.
- Teeter KC, Thibodeau LM, Gompert Z, Buerkle AC, Nachman MW, Tucker P. 2010. The variable genomic architecture of isolation between hybridizing species of house mice. *Evolution.* 64:472–485.
- The Heliconius Genome Consortium. 2012. Butterfly genome reveals promiscuous exchange of mimicry adaptations among species. *Nature.* 487:94–98.
- Turner TL, Hahn MW. 2007. Locus- and population-specific selection and differentiation between incipient species of *Anopheles gambiae*. *Mol Biol Evol.* 24:2132–2138.
- Turner TL, Hahn MW, Nuzhdin SV. 2005. Genomic islands of speciation in *Anopheles gambiae*. *PLoS Biol.* 3:e285.
- Vernot B, Akey JM. 2014. Resurrecting surviving Neandertal lineages from modern human genomes. *Science.* 343:1017–1021.
- Via S. 2009. Natural selection in action during speciation. *Proc Natl Acad Sci USA.* 106:9939–9946.
- Via S. 2012. Divergence hitchhiking and the spread of genomic isolation during ecological speciation-with-gene-flow. *Philos Trans R Soc B.* 367:451–460.
- Via S, Conte G, Mason Foley C, Mills K. 2012. Localizing FST outliers on a QTL map reveals evidence for large genomic regions of reduced gene exchange during speciation-with-gene-flow. *Mol Ecol.* 21:5546–5560.
- Via S, Hawthorne DJ. 2001. Genetic linkage of ecological specialization and reproductive isolation in pea aphids. *Nature.* 412:904–907.
- Via S, West J. 2008. The genetic mosaic suggests a new role for hitchhiking in ecological speciation. *Mol Ecol.* 17:4334–4345.
- Wagner CE, Keller I, Wittwer S, Selz OM, Mwaiko S, Greuter L, Sivasundar A, Seehausen O. 2013. Genome-wide RAD sequence data provide unprecedented resolution of species boundaries and relationships in the Lake Victoria cichlid adaptive radiation. *Mol Ecol.* 22:787–798.

- Weetman D, Wilding C, Steen K, Pinto J, Donnelly MJ. 2011. Gene flow-dependent genomic divergence between *Anopheles gambiae* M and S forms. *Mol Biol Evol.* 29:279–291.
- White BJ, Cheng C, Simard F, Costantini C, Besansky NJ. 2010. Genetic association of physically unlinked islands of genomic divergence in incipient species of *Anopheles gambiae*. *Mol Ecol.* 19:925–939.
- Whitney KD, Randell RA, Rieseberg LH. 2006. Adaptive introgression of herbivore resistance traits in the weedy sunflower *Helianthus annuus*. *Am Nat.* 167:794–807.
- Wilding C, Butlin RK, Grahame JW. 2001. Differential gene exchange between parapatric morphs of *Littorina saxatilis* detected using AFLP markers. *J Evol Biol.* 14:611–619.
- Wood HM, Grahame JW, Humphray S, Rogers J, Butlin RK. 2008. Sequence differentiation in regions identified by a genome scan for local adaptation. *Mol Ecol.* 17:3123–3135.
- Wu C-I. 2001. The genic view of the process of speciation. *J Evol Biol.* 14:851–865.
- Yang X, Xia H, Wang W, Wang F, Su J, Snow AA, Lu B-R. 2011. Transgenes for insect resistance reduce herbivory and enhance fecundity in advanced generations of crop-weed hybrids of rice. *Evol Appl.* 4:672–684.
- Yatabe Y, Kane NC, Scotti-Saintagne C, Rieseberg LH. 2007. Rampant gene exchange across a strong reproductive barrier between the annual sunflowers, *Helianthus annuus* and *H. petiolaris*. *Genetics.* 175:1883–1893.

Received February 13, 2014; First decision April 2, 2014;  
Accepted April 22, 2014

Corresponding Editor: Sean Mullen